

# Enzymes

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## INTRODUCTION

Enzymes are biological materials with catalytic properties, i.e. they increase the rate of chemical reactions in biological and in vitro (in the laboratory) systems, that otherwise proceed very slowly. The study of enzymes and of the changes in the enzyme activity that occur in body fluids has become a valuable diagnostic tool for the elucidation of various diseases and for testing organ function.

## DEFINITION

Enzymes are biological catalyst produced by living tissues. They are proteins (except a small group of RNA acting as ribozyme) that have the property of accelerating specific chemical reactions without being consumed in the process.

## ZYMOGEN OR PROENZYME

- A number of proteolytic enzymes found in the blood or in the digestive tract are present in an inactive (precursor) form, called zymogen or proenzymes.
- For example, chymotrypsin is secreted by the pancreas as chymotrypsinogen. It is activated in the digestive tract by the proteolytic enzyme trypsin.

Precursor proteins or inactive enzyme names have the prefix "pro" like prothrombin, proelastase, etc. or suffix "ogen" like chymotrypsinogen, trypsinogen, pepsinogen, etc.

## COFACTORS (COENZYME AND ACTIVATOR)

- Some enzymes require an additional nonprotein component for its optimum activity. This additional component is called cofactor which may be either loosely or tightly bound to the protein portion of the enzyme.
- These cofactors may be:
  - Organic compounds, called coenzymes
  - Inorganic ions, called activators.
- Enzymes without its cofactor is referred to as an apoenzyme; the complete catalytically active enzyme is called holoenzyme.

Apoenzyme + cofactor = holoenzyme.

- Many vitamins function as coenzymes. Coenzymes derived from vitamins will be considered for more details in chapter 7.
- The lists of coenzyme and activators are given in Tables 6.1 and 6.2 respectively.

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Table 6.1: Some common coenzymes and their functions			
Vitamin	Coenzyme	Function as coenzyme	
Thiamine (Vit B <sub>1</sub> )	TPP (Thiamine pyrophosphate)	Oxidative decarboxylation and transketolase reaction	
Riboflavin (Vit B <sub>2</sub> )	FAD and FMN (Flavin Adenine Dinucleotide and Flavin Mononucleotide)	Oxidation and reduction reactions	
Niacin (B3)	NAD* (Nicotinamide Adenine Dinucleotide), NADP* (Nicotinamide Adenine Dinucleotide Phosphate)	Oxidation and reduction reactions	
Pyridoxine (Vit B <sub>6</sub> )	PLP (Pyridoxal phosphate)	Transamination, deamination decarboxylation reactions of amino acids	
Diation ( B1)	Biocytin	Carboxylation reactions	
Form Coly (114 Ba)	Biocytin THF (Tetrahydrofolate)	Carrier of one carbon group	
Pantothenic acid (B	C) Coenzyme A	Acyl carrier	
Cynocobalamine (B)	Methylcobalamine and Deoxyadenosylcobalamine	Transfer of CH <sub>3</sub> group and isomerizations	

Enzyme	Cofactor (activator)
Ferroxidase (ceruloplasmin), Ascorbic acid oxidase	Copper
Carbonic anhydrase, DNA-polymerase, Porphobilinogen synthase, Carboxypeptidase	Zinc
Cytochrome oxidase, Catalase	Iron
Glucose-6-Phosphatase, Hexokinase	Magnesium
Glutathione peroxidase	Selenium
Arginase, Pyruvate carboxylase	Manganese
Xanthine oxidase	Molybdenum

## **HOW ENZYMES WORK**

## **Energy Changes Occur During the Reaction**

Virtually all chemical reactions have an energy barrier, separating the reactants and the products. This barrier, called the *free energy of activation*, is the energy difference between the energy of the reactant and high energy intermediates that occurs during the formation of a product. Figure 6.1 shows the changes in energy during the conversion of a molecule of reactant 'S' to product 'P' through the transition state S\*.

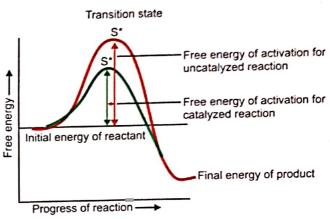
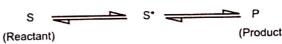


Figure 6.1: Comparison of the free energy of activation of a catalyzed and uncatalyzed reaction, S\*: Transition state



The peak of free energy activation, represents the transition state, in which the high energy intermediates (S\*) are formed during the conversion of a reactant to a product (Figure 6.1).

- An enzyme lowers the energy required for activation to the transition state.
- Without a catalyst, the reaction will occur only if enough heat energy is added to the reaction system.
- With an enzyme as a catalyst, the reaction may easily proceed at the normal physiological temperature.

## MECHANISM OF ENZYME ACTION

 Formation of an enzyme-substrate (ES) complex is the first step in enzymatic catalysis. Substrate is bound through multiple noncovalent interactions at the active site of the enzyme forming an enzyme-substrate (ES) complex which is subsequently converted to product and free enzyme.

- The active site of an enzyme is the region that binds the substrate and which contains the specific amino acid residues.
- Two models for substrate binding to the active site of the enzyme, have been proposed to explain the specificity that an enzyme has for its substrate:
  - Lock and key model or rigid template model of Emil Fisher.
  - Induced fit model or hand-in-glove model of Daniel E Koshland.

## Lock and Key Model or Rigid Template Model of Emil Fisher

- This model was proposed by Emil Fisher in 1890.
- In this model, enzyme is preshaped and the active site has a rigid structure that is complementary to that of the substrate (Figure 6.2).
- This model is called lock and key model, because in this model the substrate fits into the active site in much the same way that a key fits into a lock.
- This model has been useful in understanding how some enzymes can bind only a specific substrate but will not bind another compound with an almost identical structure. For example, most enzymes in carbohydrate metabolism can bind the D-isomer of hexoses but cannot bind the corresponding L-isomer, which differs only in the configuration around a single carbon atom.
- This model explains all mechanisms but do not explain the changes in the enzyme activity in the presence of allosteric modulators.

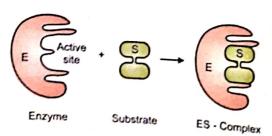


Figure 6.2: Representation of formation of an ES-complex according to the Fisher's lock and key model. The active site of the enzyme is complementary in shape to that of substrate

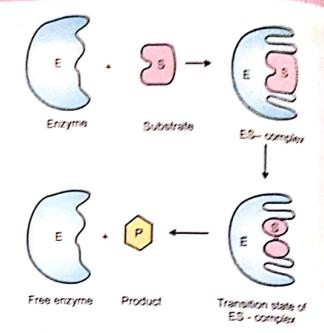


Figure 6.3: Schematic representation of induced fit model of Koshland

# Induced Fit Model or Hand-in-glove Model of Daniel E Koshland (Figure 6.3)

Fisher's model explained the specificity of enzyme substrate interaction but the implied rigidity of the enzymes active site failed to explain the dynamic changes that must take place during catalysis. A model that accounts for both of these aspects of enzyme catalysis is the induced fit model of Daniel E Koshland.

- In the Fisher's model the catalytic site is presumed to be preshaped to fit the substrate. However, Daniel E Koshland in 1958 postulated that the enzymes are modified by the binding of the substrate.
- In the induced fit model, the substrate induces a conformational change in the enzyme, in the same manner in which placing a hand (substrate) into a glove (enzyme) induces changes in the glove's shape. Therefore, this model is also known as hand-in-
- Conformational change in enzyme in turn induces reciprocal changes in its bound substrate that alters their orientation and configuration and strains the structure of the bound substrate. Due to such changes energy is librated, which is called intrinsic binding energy (Figure 6.3).
- This intrinsic binding energy due to the substrateenzyme interaction is made available for the transformation of the substrate into product.

This model is believed to describe more accurately the specificity of substrate binding than does lock and key model of E Fisher.

## **ENZYME CLASSIFICATION**

The classification of enzyme was described in 1961 by enzyme commission of the International Union of Biochemistry (IUB). According to this classification, each enzyme is characterized by a code number called enzyme code number or 'EC' number, consisting of four digits.

According to the IUB system, enzymes are classified into six major classes as follows:

1. EC-1: Oxidoreductases

2. EC-2: Transferases

3. EC-3: Hydrolases

4. EC-4: Lyases

5. EC-5 : Isomerases

6. EC-6: Ligases.

Some common examples of different classes of enzymes are given in Table 6.3.

## **EC-1 Oxidoreductases**

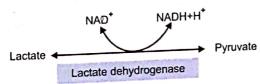
Those enzymes that catalyze oxidation-reduction reactions, are included in this class which can be illustrated schematically as follows:

$$A + B \longrightarrow A + B$$
(red) (ox) (red)

Enzymes in this category include:

- Dehydrogenases
- Reductases
- Oxidases
- Peroxidases.

## Specific Example



## **EC-2 Transferases**

Those enzymes that catalyze the transfer of a group such as, amino, carboxyl, methyl or phosphoryl, etc. from one molecule to another are called transferases. These reactions can be illustrated as follows:

Table 6.3: Examples	of enzymes with their main class
Class	Example
EC-1:Oxidoreductases	Lactate dehydrogenase (LDH) Glucose 6-phosphate dehydrogenase (G-6-PD) Cytochrome oxidase
EC-2: Transferases	Aspartate aminotransaminase(AST Alanine aminotransaminase (ALT) Hexokinase
EC-3: Hydrolases	Lipase α-Amylase Trypsin Lactase Sucrase
	Pepsin
EC-4: Lyases	Aldolase Argininosuccinase Carbonic anhydrase
EC-5: Isomerases	Phosphoglucomutase Triphosphate isomerase Phosphohexose isomerase
EC-6: Ligases	Glutamine synthetase Pyruvate carboxylase DNA ligases

Some common enzymes in this category include:

- Amino transferase or transaminase
- Kinase
- Transcarboxylase.

#### Specific Example

Alanine + 
$$\alpha$$
-Ketoglutarate  $\alpha$ -Ketoglutarate  $\alpha$ -Ketoglutarate  $\alpha$ -PLP

ALT: Alanine aminotransferase where,

PLP: Pyridoxal phosphate (coenzyme)

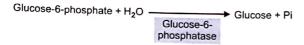
## **EC-3 Hydrolases**

Enzymes of this class catalyze the cleavage of C-O, C-N, C-C and some other bonds with the addition of water. These can be illustrated as follows:

Some common enzymes in this category are:

- Acid phosphatase
- All digestive enzymes like  $\alpha$ -amylase, pepsin, trypsin, chymotrypsin, etc.

### Specific Example



#### **EC-4 Lyases**

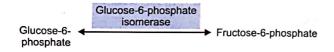
Lyases catalyze the cleavage of C-O, C-C and C-N bonds by means other than hydrolysis or oxidation, giving rise to compound with double bonds or catalyze the reverse reaction, by the addition of group to a double bond. In cases where reverse reaction is important, then synthase, (not synthetase of group EC-6) is used in the name. This type of reaction is illustrated as follows:

### Specific Example

#### **EC-5** Isomerases

Isomerases catalyze intramolecular structural rearrange ment in a molecule. They are called epimerases, isomerases or mutases, depending on the type of isomerism involved. This reaction can be illustrated as follows:

### Specific Example



## EC-6 Ligases (Synthetases)

Ligases catalyze the joining of two molecules coupled with the hydrolysis of ATP. The reaction is illustrated as follows:

#### Specific Example

## SPECIFICITY OF ENZYME ACTION

Specificity refers to the ability of an enzyme to discriminate between two competing substrates. Enzymes are highly specific both in the reaction catalyzed and in their choice of substrates. Specificity makes it possible for a number of enzymes to co-exist in the cell without interfering in each other's actions.

## Types of Specificity

The following types of specificity have been recognized:

- 1. Substrate specificity
- 2. Reaction specificity
- 3. Stereo specificity.

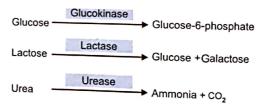
### Substrate Specificity

There are following types of substrate specificity:

- i. Absolute substrate specificity
- ii. Relative substrate specificity
- iii. Broad substrate specificity.

## Absolute substrate specificity

Certain enzymes will act on only one substrate and catalyze one reaction, e.g. Glucokinase, lactase, urease, etc.



## Relative substrate specificity

In this case, enzyme acts on more than one substrate. The relative substrate specificity is of two types:

- a. Group specificity
- b. Bond specificity.
- In group specificity, an enzyme acts on more than one substrate containing a particular group, e.g. chymotrypsin acts on several proteins by hydrolyzing peptide bonds attached to aromatic amino acids. Trypsin hydrolyzes peptide linkages involving arginine or lysine.
- In bond specificity, an enzyme acts on more than one substrate containing a particular kind of bond, e.g. salivary  $\alpha$ -amylase cleaves  $\alpha$ -(1 $\rightarrow$ 4) glycosidic bonds of carbohydrates, lipase hydrolyzes ester bonds of lipids.

## Broad substrate specificity

In this case, an enzyme acts on more than one structurally related substrates, e.g. hexokinase catalyzes the phosphorylation of more than one kind of hexoses such as glucose, fructose and mannose.

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#### Reaction Specificity

In this case, an enzyme is specific to a particular reaction but not to substrate (s) and catalyzes only one type of reaction. For example, pyruvate can undergo several reactions. Each reaction is catalyzed by a separate enzyme, which catalyzes only that reaction and none other as shown in Figure 6.4.

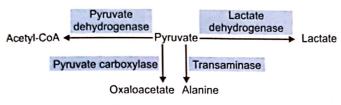


Figure 6.4: Example of reaction specificity

#### Stereo Specificity

Many enzymes show specificity towards stereoisomers, i.e. they act on only one type of isomer. For example,

- L-lactate dehydrogenase will act only on L-lactic acid and not D-lactic acid.
- Likewise, L-amino acid oxidase and D-amino acid oxidase are distinct enzymes which act only on L and D-amino acids respectively.
- D-glucose oxidase can similarly act only on Dglucose and not on L-glucose.
- Salivary α-amylase acts on the α-1,4 glycoside linkage and is inactive on β-1,4 glycoside linkage.
- Isomerase and epimerase do not show stereospecificity.

## FACTORS AFFECTING THE VELOCITY OF ENZYME REACTION

Various factors that affect enzyme activity are:

- Substrate concentration
- Enzyme concentration
- pH i.e. H<sup>1</sup> ion concentration
- Temperature
- Product concentration
- Activators and coenzymes
- Time
- Physical agents
- Inhibitors

#### **Effect of Substrate Concentration**

For a given quantity of enzyme, the velocity of the reaction increases as the concentration of the substrate is increased. At first, this relationship is almost linear but later, the reaction curve becomes hyperbolic in shape (Figure 6.5).

At relatively low concentrations of substrate, V<sub>0</sub> increases almost linearly with an increase in substrate concentration [S], a condition known as First order kinetics.

- At higher substrate concentrations, V<sub>0</sub> increases by smaller amounts in response to increase in substrate concentration [S].
- Finally, a part is reached beyond which there are only vanishingly small increase in V<sub>0</sub> with increasing [S], a condition known as *zero order kinetics* and a plateau is called *maximum velocity*, V<sub>max</sub> (Figure 6.5). Under these conditions, all the free enzymes will have been converted into ES form so that any further increase in substrate concentration has no effect on the rate and the reaction achieves a steady state.

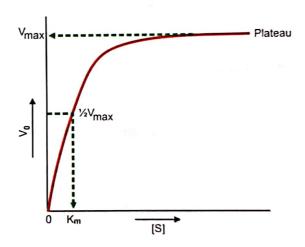


Figure 6.5: Effects of substrate concentration [S] on enzyme activity keeping enzyme concentration constant

where, V<sub>0</sub>: initial velocity

V<sub>max</sub>: maximum velocity

K<sub>m</sub>: 1/2 V<sub>max</sub> = Michaelis Menten constant

[S] : substrate concentration

#### **Effect of Enzyme Concentration**

- The velocity of a reaction is directly proportional to the amount of enzyme present as long as the amount of substrate is not limiting.
- The substrate must be present at a concentration sufficient to ensure that all of the enzyme molecules have substrate bound to their active site (Figure 6.6).

#### Effect of Hydrogen Ion Concentration pH

 Each enzyme has an optimum pH, i.e. a pH at which the enzyme activity is maximum. Below or above this pH, enzyme activity is decreased. The optimum pH

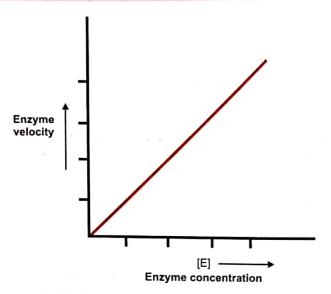


Figure 6.6: Effect of enzyme concentration on enzyme activity

differs from enzyme to enzyme, e.g. optimum pH for:

- Pepsin = 1.2
- Trypsin = 8.0
- A bell shaped curve is obtained when enzyme velocity is plotted against pH (Figure 6.7).
- Changes in pH can alter the following:
  - Ionization state of the amino acid residues present in the active site of the enzyme. Enzyme activity is related to the ionic state of active site of the enzyme.
  - The ionization state of the substrate. The active site of an enzyme may require particular ionic state of the substrate for optimum activity.
  - Drastic change in pH denatures the enzyme protein.

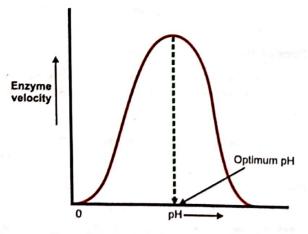


Figure 6.7: Effect of pH on enzyme activity

All these affect the enzyme substrate complex formation and decrease the rate of enzyme reaction. As pH affects the enzyme activity, in enzyme studies, buffers are used to keep enzyme at an optimum or at least a favorable H<sup>+</sup> ion concentration.

## **Effect of Temperature**

- Enzyme catalyzed reactions show an increase in rate with increasing temperature only within a relatively small and low temperature range.
- Each enzyme shows the highest activity at a particular temperature called optimum temperature.
- The activity progressively declines both above and below this temperature.
- A bell shaped curve is obtained when we plot the enzyme velocity Vs. temperature (Figure 6.8).

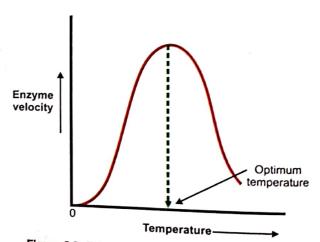


Figure 6.8: Effect of temperature on enzyme activity

- Increase in velocity is due to the increase in the kinetic energy. Further elevation of the temperature results in a decrease in reaction velocity due to denaturation of the enzyme protein.
- Low temperature also decreases enzyme activity and enzymes may be completely inactive at temperature of 0°C and below.
- The inactivity at low temperature is reversible. So, many enzymes in tissues or extracts may be preserved for months by stories.
- preserved for months by storing at -20°C or -70°C.
   The reaction velocity of most chemical reactions increases with temperature approximately doubles for each 10°C rise called temperature.
- for each 10°C rise called temperature coefficient Q<sub>10</sub>.

   Most of the body enzymes have the optimum temperature close to 37°C to 38°C and have progressively less activity as the temperature rises.

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## **Effect of Product**

Accumulation of products of the reaction causes the inhibition of enzyme activity for some enzymatic reactions, this form of control will limit the rate of formation of the product when the product is under used. In biological systems, however, the product is usually removed as it becomes a substrate for a succeeding enzyme in a metabolic pathway.

## Effect of Activators and Co-enzymes

The activity of many enzymes is dependent on the activators (metallic ions) like Mg<sup>2+</sup>, Mn<sup>2+</sup>, Zn<sup>2+</sup>, Ca<sup>2+</sup>, Co<sup>2+</sup>, Cu<sup>2+</sup>, etc. and coenzymes for their optimum activity. In absence of these activators and coenzymes, enzymes become functionally inactive.

## **Effect of Time**

Under optimum conditions of pH and temperature, time required for an enzyme reaction is less. The time required for the completion of an enzyme reaction increases with changes in temperature and pH from its optimum.

## **Effect of Physical Agents**

Physical agent like light rays can inhibit or accelerate certain enzyme reactions. For example, the activity of salivary amylase is increased by red and blue light. On the other hand, it is inhibited by ultraviolet rays.

## Effect of Inhibitors

The substances which stop the enzymatic reaction are called inhibitors. Presence of these substances in reaction medium decreases the rate of enzyme reaction. Different types of enzyme inhibitors are discussed separately later.

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## **ENZYME KINETICS**

The study of enzyme reaction rates and how they change in response to changes in experimental parameters is known as *kinetics*.

One of the key factors affecting the rate of a reaction catalyzed by an enzyme *in vitro* (in laboratory) is the amount of substrate present, [S]. The effect on V<sub>0</sub> (initial velocity) of varying substrate [S] concentration, when enzyme concentration is held constant, is shown in Figure 6.5.

## Michaelis-Menten Equation

Enzyme catalyzed reactions occur in two stages as shown in the following equations

E+S 
$$\frac{K_1}{K_2}$$
 ES.....1)

 $K_1$ ,  $K_2$  and  $K_3$  are rate constant. The Michaelis-Menten equation describes how reaction velocity varies with substrate concentration. The relationship between reaction rate and substrate concentration shown in **Figure 6.5** is described mathematically by the 'Michaelis-Menten equation' as follows:

$$V_0 = \frac{V_{\text{max}} [S]}{K_{\text{m}} + [S]}$$

where,

 $V_0$  = initial reaction velocity (is the rate of reaction as soon as enzyme and substrates are mixed).

V<sub>max</sub> ≈ maximum velocity (is observed when all active sites on the enzyme are filled with substrate).

 $K_m$  = Michaelis-Menten constant, [is the substrate concentration, at which the reaction rate is half of its maximum velocity  $(V_{max})$ ].

[S] = Substrate concentration

Every enzyme has the characteristics  $V_{\text{max}}$  and  $K_{\text{m}}$  which are sensitive to change in pH, temperature and ionic strength.

## Significance of $K_m$ (Michaelis Constant)

- K<sub>m</sub>, the Michaelis constant is equal to the substrate concentration at which the reaction rate is half of its maximal value. The K<sub>m</sub> value of an enzyme depends on the substrate and environmental conditions such as pH, temperature and ionic strength.
- The Michaelis constant, K<sub>m</sub> has two significances:
  - K<sub>m</sub> provides a measure of the substrate concentration required for significant catalysis to occur.
  - 2. It is a measure of the affinity of the enzyme for its substrate, a high  $K_m$  indicates weak binding and a low  $K_m$  indicates strong binding with its substrate.

## Significance of $V_{max}$ (Maximal Velocity)

• The  $V_{max}$  of a reaction is an index of the catalytic efficiency of an enzyme. The  $V_{max}$  is useful in comparing the activity of one enzyme with that of another.

## Lineweaver-Burk Plot or Double-Reciprocal Plot

Lineweaver-Burk plots are used to obtain values for  $V_{\text{max}}$  and  $K_{\text{m}}$ . A more accurate method of determining

values for  $V_{max}$  and  $K_m$  uses Lineweaver-Burk equation shown below. This equation is obtained by taking the reciprocal of the Michaelis-Menten equation.

$$\frac{1}{V_0} = \frac{K_m}{V_{\text{max}}} \quad \frac{1}{[S]} + \frac{1}{V_{\text{max}}}$$

- When 1/V<sub>0</sub> is plotted against 1/[S], a straight line is obtained (Figure 6.9), with a slope equal to K<sub>m</sub>/V<sub>max</sub>.
- The point, at which line intersects the y-axis, is numerically equal to 1/V<sub>max</sub>.
- The point at which the line intersects the x-axis is numerically equal to -1/K<sub>m</sub>.
- This plot is useful to determine the mechanism of action of enzyme inhibitors.

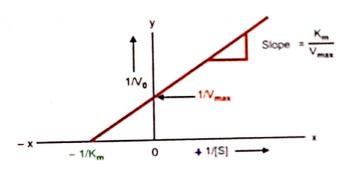


Figure 6.9: Lineweaver-Burk plot (Double reciprocal plot)

## **ENZYME INHIBITION**

Any substance that can diminish the velocity of an enzyme catalyzed reaction is called inhibitor. Two general classes of inhibitors are recognized according to whether the inhibitor action is reversible or irreversible (Figure 6.10)

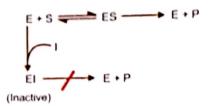
- Reversible inhibitor
- 2. Irreversible inhibitor.

## Reversible Inhibitor

- Reversible inhibitors bind to enzymes through noncovalent bonds and the activity of the enzyme is restored fully when the inhibitor is removed from the system.
- Different types of reversible inhibitors are:
  - i. Competitive or substrate analogue inhibitor
  - ii. Noncompetitive inhibitor
  - iii. Uncompetitive inhibitor.

## Competitive or Substrate Analogue Inhibitor

- A competitive inhibitor is usually a structural analogue of the substrate.
- The chemical structure of the inhibitor (I) closely resembles that of the substrate (S) and binds to the enzyme at the active site, forming an EI complex rather than ES-complex (Figure 6.11).



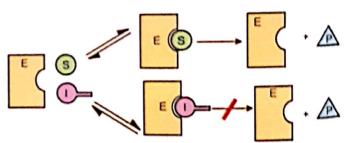


Figure 6.11: Diagrammatic representation of competitive inhibition where, E: Enzyme; S: Substrate; I: Competitive inhibitor; P: Product

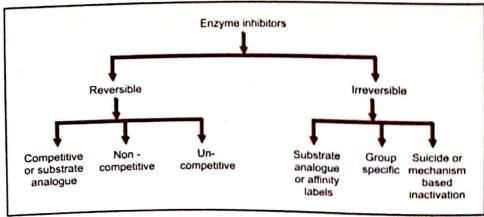


Figure 6.10: Classes of enzyme inhibitors

- But because it is not identical with the substrate. breakdown into products does not take place.
- When both the substrate and this type of inhibitor are present, they compete for the same binding site on the enzyme.
- The inhibition could be overcome by increasing substrate concentration. In competitive inhibition, the K<sub>m</sub> increases whereas V<sub>max</sub> remains unchanged (Figure 6.12).

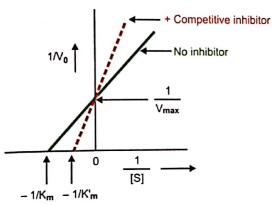


Figure 6.12: A double reciprocal plot of enzyme kinetics in presence and absence of competitive inhibitor,  $V_{\text{max}} \, \text{is}$  unaltered whereas  $K_{\text{m}} \, \text{is}$ increased

- The classical example is the competitive inhibition of succinate dehydrogenase by the malonate.
- Succinate dehydrogenase is one of the enzymes of the citric acid cycle. Succinate dehydrogenase is inhibited by malonate which resembles succinate.
- Many drugs which act as competitive inhibitors are given below and some more are given in Table 6.4.

#### Sulfonamide

Sulfonamide is an analogue of P-aminobenzoic acid (PABA) and inhibits pteroid synthetase enzyme required for the synthesis of folic acid in microorganisms.

Since folic acid is involved in the biosynthesis of purines and thymine, sulfonamides inhibit growth of the pathogenic organisms.

## Isoniazide [Isonicotinic acid hydrazine (INH)]

It is an antituberculous drug, an analogue of nicotinamide.

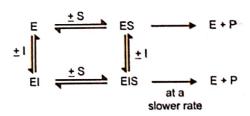
INH interferes with the biosynthesis of NAD and restrict the growth of the organisms that cause tuberculosis.

### Dicumarol

It is an anticoagulant drug structurally similar to vitamin K. It inhibits the vitamin K activity and inhibits the formation of prothrombin.

## Noncompetitive Inhibitors

- As the name implies, in this type of inhibition no competition occurs between substrate and inhibitor. Inhibitor is usually structurally different from the substrate.
- It binds at a site on the enzyme molecule other than the substrate-binding site and thus there is no competition between inhibitor and substrate.
- Since inhibitor and substrate may bind at different sites; formation of both EI and EIS-complexes is possible.
- Since EIS may breakdown to form product at a slower rate than does ES, the reaction is slowed but not halted. The following reactions may occur:



For noncompetitive inhibition, the K<sub>m</sub> value is unchanged while V<sub>max</sub> is lowered (Figure 6.13)

Dava	Type of Inhibition	Target enzyme	Therapeutic use
Drug Mevinolin Lovastatin	Competitive	HMG-CoA reductase (3-Hydroxy-3-Methyl-Glutaryl CoA-reductase)	Hypercholesterolemia
Allopurinol Methotrexate 5-Fluorouracil	Competitive Competitive Suicide Suicide	Xanthine oxidase  Dihydrofolate reductase  Thymidylate synthase  Cyclo-oxygenase	Gout Cancer Cancer Anti-inflammatory
Aspirin Penicillin	0.1-140	Bacterial transpeptidase	Antibacterial

- Examples of noncompetitive inhibitors are:
- Ethanol or certain narcotic drugs noncompetitive inhibitor of acid phosphatase.
- Trypsin inhibitors occur in soybean and raw egg white, inhibit activity of trypsin noncompetitively.
- As well as Ascaris parasites (worm) contain pepsin and trypsin inhibitors, which inhibit noncompetitively action of pepsin and trypsin, that is why ascaris worm is not digested in human intestine.

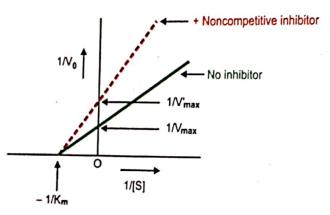


Figure 6.13: A double reciprocal plot of enzyme kinetics in presence and absence of noncompetitive inhibitor;  $K_{\text{m}}$  is unaltered by noncompetitive inhibitor, whereas  $V_{\text{max}}$  is decreased

## Uncompetitive Inhibitor

- Uncompetitive inhibitor can bind only to the enzyme-substrate (ES) complex.
- It does not have affinity for free enzyme. Uncompetitive inhibitor decreases both  $V_{\text{max}}$  and Km (Figure 6.14).
- This form of inhibitor is rare with single substrate but more common in multiple substrate reaction.

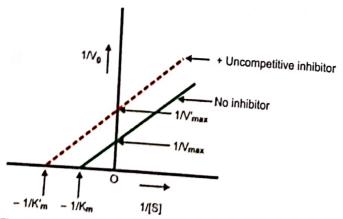


Figure 6.14: Effect of uncompetitive inhibitor on the double reciprocal plot; it shows parallel lines with decrease in both  $V_{\text{max}}$  and  $K_{\text{m}}$ 

## Irreversible Inhibitor

- An irreversible inhibitor binds with an enzyme tightly covalently and forms a stable complex. An irreversible inhibitor cannot be released by
- dilution or dialysis or simply by increasing the concentration of substrate.
- Irreversible inhibitors can be divided into three categories:
  - i. Group specific inhibitors
  - ii. Substrate analogue inhibitor or affinity labels
  - iii. Suicide inhibitor or mechanism based inactivation. In terms of enzyme kinetics, the effect of an
- irreversible inhibitor is like that of the reversible noncompetitive inhibitors resulting in a decreased in  $V_{max}$  but having no effect on the  $K_m$  (Table 6.5).

Table 6.5: Effect of properties	s of enzymes	i kinetic
Type of inhibitor	Km	V <sub>max</sub>
Irreversible	No effect	Decreased
Reversible competitive	Increased	No effect
Reversible noncompetitive	No effect	Decreased
Reversibe uncompetitive	Decreased	Decreased

## Group Specific Irreversible Inhibitor

These inhibitors react with specific R-groups (side chain) of amino acid residues in the active site of enzyme. Examples of group specific irreversible inhibitors are:

# Di-isopropylphosphofluoride (DIPF)

DIPF can inhibit an enzyme acetylcholine esterase by covalently reacting with hydroxyl group of a serine residue present at the active site of the enzyme

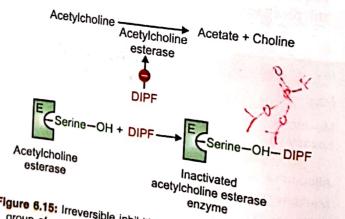


Figure 6.15: Irreversible inhibition of acetylcholine esterase by a group of specific inhibitor, diisopropylphosphofluoride (DIPF)

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 DIPF has also been found to inhibit trypsin, chymotrypsin, elastase and phosphoglucomutase.

Iodoacetamide and heavy metals

Iodoacetamide and heavy metals like, Pb<sup>2+</sup>, Ag<sup>+</sup>, Hg<sup>2+</sup>, etc. which react with sulfhydryl (-SH) group of cysteine residues present at the active site of the enzyme and makes them inactive.

Substrate Analogue Irreversible Inhibitor or Affinity Labels

- Substrate analogues or affinity labels are molecules that are structurally similar to the substrate.
- These substrate analogues possess a highly reactive group which is not present in the natural substrate.
- The reactive group of substrate analogues covalently reacts with amino acid residues of the active site of the enzyme and permanently block the active site of the enzyme, e.g. 3-Bromoacetol phosphate (BAP).
- BAP is a substrate analogue of the normal substrate dihydroxyacetone phosphate (DHAP) for the enzyme phosphotriose isomerase of glycolysis.

Suicide Inhibitor or Mechanism Based Inactivation

- These compounds are relatively unreactive until they bind to the active site of a specific enzyme.
- On binding to the active site of the enzyme they carry out the first few catalytic activities of the normal enzyme reaction.
- Instead of being transformed into a normal product, however, the inhibitor is converted to a very reactive compound that combines irreversibly with the enzyme leading to its irreversible inhibition.
- The enzyme literally commits suicide. These are also called mechanism based inactivation because they utilize the normal enzyme reaction mechanism to inactivate the enzyme.
- These inhibitors act as drugs for example.

Penicillin

 Penicillin irreversibly inactivates an essential bacterial enzyme glycopeptidyl transpeptidase involved in the formation of bacterial cell wall.

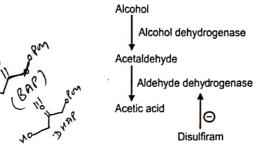
Aspirin

 Aspirin inactivates an enzyme cyclo-oxygenase which catalyzes the first reaction in the biosynthesis of prostaglandins from arachidonic acid.

Disulfiram (antabuse)

Disulfiram is a drug used in the treatment of alcoholism. It inhibits irreversibly aldehyde dehydro-

genase enzyme resulting in accumulation of acetaldehyde in the body. Accumulation of acetaldehyde in the tissue leads to alcohol avoidance.



## Clinical Application of Enzyme Inhibitor

Enzyme inhibitors have therapeutic applications. Most antibiotics and anticancer drugs that are used therapeutically are either competitive inhibitor or mechanism based suicide inhibitor. Some commonly used drugs that exert their effects by inhibiting enzymes are described in **Table 6.4.** 

#### ALLOSTERIC ENZYME

Allosteric enzyme is a regulatory enzyme. The term allosteric derives from Greek word, allo means other and steros means space or site. Allosteric enzymes are those having other site.

Like all enzymes, allosteric enzymes have active site for binding of the substrate but they also have one or more regulatory (or allosteric) sites for binding regulatory metabolites which is called *modulator*.

- Allosteric enzymes may be inhibited or stimulated by their modulators (Table 6.6).
- Modulators that inhibit enzyme activity are termed negative modulators. Whereas those that increase enzyme activity are called positive modulators.
- Just as the active site of an enzyme is specific for its substrate, the allosteric site is specific for its modulators.
   Allosteric enzymes are generally larger and more complex than those of simple enzymes.

#### Feedback Allosteric Inhibition

In some multienzyme systems, the first enzyme of the sequence is the regulatory enzyme and has distinctive characteristics.

 It is inhibited by the end product of the multienzyme system whenever the end product of such metabolic reaction produced in excess of the cells needs. The end product of the pathway acts as a specific inhibitor of the first or regulatory enzyme in the pathway.

	Table 6.6: Allosteric enzymes and		Activator
Pathway	Enzyme	Inhibitor	
	Phosphofructokinase-I	ATP	AMP
Glycolysis	Pyruvate dehydrogenase	ATP	
Pyruvate to acetyl-CoA	Isocitrate dehydrogenase	ATP	ADP
TCA cycle		•	Acetyl-CoA
Gluconeogenesis Fatty acid synthesis	Pyruvate carboxylase Acetyl-CoA carboxylase		Citrate

 The whole enzyme system thus slows down to bring the rate of production of its end product back into balance with the cell's needs.

This type of regulation is called *feedback inhibition*. For example, first enzyme  $\delta$ -aminolevulenic acid synthase is an allosteric enzyme in heme synthesis is inhibited by its end product heme. (Figure 6.16).

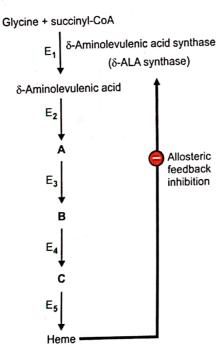


Figure 6.16: Feedback inhibition. First enzyme  $\delta$ -ALA-synthase is an allosteric enzyme inhibited by its end product heme

#### ISOENZYME

Isoenzymes or isozymes are multiple forms (isomers)
of the same enzyme that catalyze the same
biochemical reaction. Isoenzymes show different
chemical and physical properties like electrophoretic
mobility and kinetic properties.

- Not all enzymes have isoenzymes. In fact, it was found that only those enzymes, which are active in polymeric form demonstrate isoenzyme. For example:
  - 1. Lactate dehydrogenase (LDH)
  - Creatine kinase (CK) (formerly called creatine phosphokinase (CPK).
- Some more examples of isoenzymes are given in Table 6.7.

Table 6.7: Examples of isoenzymes			
Enzyme	Isoenzyme forms		
Acid phosphatase	Prostate, erythrocytes, platelets, liver, spleen, kidney and bone marrow		
Alkaline phosphatase	Bone, liver, placenta, intestine and kidney		
Amylase	Salivary and pancreatic		
Hexokinase	Liver (glucokinase) and muscle		

## Lactate Dehydrogenase (LDH)

Lactate dehydrogenase is a tetrameric enzyme that catalyzes the oxidation of L-lactate to pyruvate.

- LDH has five isoenzymes:
  - LDH1 (Hy)
  - LDH2 (mm)
  - LDH3 (H2M2)
  - LDH4 (川My)
  - LDH<sub>5</sub>. (M<sub>4</sub>)
- Since LDH is a tetramer, made up of two types of polypeptide M (muscle) type and H (heart) type, five combinations are possible with varying ratios of two kinds of polypeptides (Table 6.8).
- Five isoenzymes of LDH can be detected by electrophoresis as they have different electrophoretic mobilities.
- LDH<sub>1</sub> is the fastest moving fraction towards the anode and LDH<sub>5</sub> is the slowest moving isoenzyme of LDH (Figure 6.17).

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 LDH<sub>1</sub> predominates in cells of cardiac muscle, and erythrocytes and LDH<sub>5</sub> is the most abundant form in the liver and in skeletal muscle (Table 6.8).

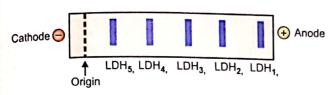


Figure 6.17: Electrophoretic separation of LDH isoenzyme

## Clinical Applications of LDH

The LDH isoenzyme analysis may be useful in the following clinical situations (Table 6.8):

- Significant elevation of LDH<sub>1</sub> and LDH<sub>2</sub> (LDH<sub>1</sub>>LDH<sub>2</sub>) occurs within 24 to 48 hours after myocardial infarction.
- Predominant elevation of LDH<sub>2</sub> and LDH<sub>3</sub> occur in leukemia. LDH<sub>3</sub> is the main isoenzyme elevated due to malignancy of many tissues.
- Elevation of LDH<sub>5</sub> occurs after damage to the liver or skeletal muscle.

### Creatine Kinase (CK)

- Creatine kinase isoenzymes are dimer that are made up of two types of polypeptide chains, which may be either *M* (*muscle*) type or *B* (*brain*) type, generating three isoenzymes (Table 6.8).
  - $-CK_1$  (BB) : It is present in the brain
  - $-CK_2$  (MB) : Cardiac tissue is the only tissue which has the mixed MB (CK<sub>2</sub>) isoenzyme
  - -CK<sub>3</sub> (MM): It is present in skeletal muscle.

#### Clinical Application

- CK<sub>1</sub> may be elevated in neonates particularly in damaged brain or very low birth weight newborn.
- Increased level of CK<sub>2</sub> in blood is characteristic of damage of heart tissue from myocardial infarction because cardiac tissue is the only tissue which has the mixed MB (CK<sub>2</sub>) isoenzyme.
- Elevated levels of CK<sub>3</sub> in serum occur in all types of dystrophies and myopathies.

## CLINICAL SIGNIFICANCE OF ENZYMES

Certain enzymes are used:

- For the diagnosis of the disease
- As therapeutic agents
- As analytical reagents.

## Diagnostic Use of Enzymes

Enzymes are known as marker of **cellular damage** and their measurement in plasma is used in the investigation of diseases of **liver**, **heart**, **skeletal muscle**, the **biliary** tract and the **pancreas**. The enzymes that are found in plasma can be categorized into two major groups: The **plasma specific** enzyme and the **plasma nonspecific** enzyme.

- The plasma specific enzymes are present in higher concentration in plasma than in most tissues; e.g.
  - The enzymes involved in blood coagulation
  - Ferroxidase
  - Pseudocholinesterase
  - Lipoprotein lipase.
- These enzymes are clinically of interest when their concentration decreases in plasma.

Table 6.8: Type, composition, location and diagnostic importance of lactate dehydrogenase (LDH) and creatine kinase (CK) isoenzymes					
Туре	Composition	Location	Diagnostic importance (cause of elevated level)		
LDH <sub>1</sub>	нннн	Heart, RBC	Myocardial infarction		
LDH <sub>2</sub>	нннм	Heart, RBC	Megaloblastic anemia		
LDH <sub>3</sub>	ннмм	Brain	Leukemia, malignancy		
LDH <sub>4</sub>	HMMM	Lung, spleen	Pulmonary infarction		
LDH <sub>5</sub>	MMMM	Liver, muscle	Liver diseases, Muscle damage/diseases		
CK <sub>1</sub>	BB	Brain	Neurological injury		
CK <sub>2</sub>	BM	Heart	Myocardial infarction		
CK <sub>3</sub>	MM	Skeletal muscle	Muscular dystrophies and myopathies		

- The plasma nonspecific enzymes are present in very high concentration in tissues than in the plasma.
   Estimation of plasma nonspecific enzymes is very important for the diagnosis of several disease.
- The important enzymes useful for the diagnosis of specific diseases are listed in Table 6.9. A brief account of selected diagnostic enzymes is discussed.

#### Alanine transaminase (ALT)

- Alanine transaminase was known formerly as glutamate pyruvate transaminase (GPT).
- The plasma ALT normal value for adult is 10 to 40 U/L.
- ALT level is elevated in liver diseases (viral or toxic hepatitis), jaundice and cirrhosis of liver.

#### Aspartate transaminase (AST)

- It was known formerly as glutamate oxaloacetate transminase (GOT).
- The plasma AST normal value for adults is 10 to 30 U./.
- Increased AST level occurs after myocardial infarction. The plasma AST level starts increasing after 6 to 8 hours after the onset of chest pain with peak values 18 to 24 hours and the values fall to normal level by the fourth or fifth day.
- It is moderately elevated in liver disease.

### Alkaline phosphatase (ALP)

- ALP hydrolyzes organic phosphate at alkaline pH.
- Normal serum level for adults is 3-13 KA units/dl.
- It is elevated in certain bone and liver disease.

 Very high levels may be noticed in obstructive jaundice, bone diseases such as Paget's disease, rickets, osteomalacia, carcinoma of bone and hyperparathyroidism.

### Acid phosphatase (ACP)

- It hydrolyzes phosphoric acid ester at pH 5 to 6.
- Normal serum value for ACP is 0.5 to 4 KA units/dL.
- Prostatic acid phosphatase enzyme is useful for the diagnosis and prognosis of prostate cancer. ACP is therefore an important tumor marker.

#### Amylase

- It catalyzes hydrolysis of starch and glycogen.
- Normal serum value is 50-120 U/L.
- The activity of serum amylase is increased in acute pancreatitis.
- Elevated activity of amylase is also found in urine of the patient of acute pancreatitis.
- Increase in serum levels are also seen in chronic pancreatitis, mumps and obstruction of pancreatic duct.

Creatine kinase (CK) Refer isoenzyme.

Lactate dehydrogenase (LDH) Refer isoenzyme.

### Prostate specific antigen (PSA Seminogelase)

 It is a glycoprotein with mild protease activity and involved in the liquification of the seminal coagulum formed after ejaculation.

Table 6:9: Enzymes of diagnostic importance			
Enzyme	Clinical Application		
Acid phosphatase	Prostatic cancer		
Alanine aminotransferase	Liver disease (viral or toxic hepatitis), jaundice and liver cirrhosis		
Aldolase	Muscle diseases		
Alkaline phosphatase	Obstructive jaundice, bone diseases such as Paget's disease, rickets, osteomalacia, carcinoma o bone and hyperparathyroidism		
Amylase	Acute pancreatitis, mumps, obstruction in pancreatic duct		
Aspartate transaminase	Myocardial infarction, liver diseases		
Cholinesterase	Organophosphorus insecticide poisoning, hepatic parenchymal diseases		
Creatine kinase	Myocardial infarction, muscle diseases		
y-Glutamyl transferase	Hepatobiliary disease, alcoholism		
Lactate dehydrogenase	Myocardial infarction, leukemia, muscular dystrophy, hepatic diseases		
5'-Nucleotidase	Hepatitis, obstructive jaundice		
Prostate specific antigen	Prostate cancer		
Trypsin	Pancreatic disease, cystic fibrosis		

- It is produced exclusively by prostate glands.
- Elevated blood levels of PSA occur in prostate cancer.

### **Enzyme Assay in Myocardial Infarction**

The enzymes' assay, which are most helpful in the diagnosis of myocardial infarction are listed in **Table 6.10** and **Figure 6.18**.

## Therapeutic Use of Enzymes

Some enzymes are used in the treatment of some diseases of human being. For example:

- Bacterial asparginase is used in the treatment of some types of leukemia. Asparginase which hydrolyzes aspargine to aspartic acid. Aspargine is necessary for the formation of leukemic white cell.
- **Chymotrypsin:** Used for dissolving ligaments of the lens during the extraction of cataract.
- Collagenase: Used for debridement (cleaning of wound by removing dead tissue) of dermal ulcers and severe burns.
- **Fibrinolysin:** It is used in the venous thrombosis, pulmonary and arterial embolism (blood clot).
- Hyaluronidase: It is used to promote the rapid absorption of drugs injected subcutaneously. It acts by increasing tissue permeability. It is used in the treatment of traumatic or postoperative edema.
- Lysozyme (an antibiotic) found in human tears and egg white, is used in the infection of eye. It has antibacterial action, it acts on cellulose of bacteria.
- Penicillinase (bacterial enzyme) is used for the treatment of persons who are allergic to penicillin, penicillinase destroys penicillin.
- Rennin is used for the treatment of gastric achylia.
- Streptokinase: Streptokinase and urokinase are used in myocardial infarction to dissolve blood clot or purulent (containing pus) material. It causes fibrinolysis.
- Trypsin: It is a proteolytic enzyme and is used to clean the wounds by dissolving purulent material and in the treatment of acute thrombophlebitis to dissolve the blood clot.

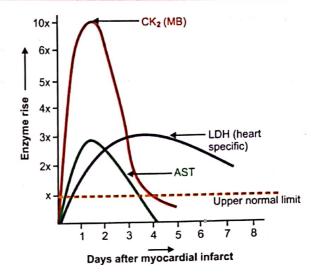


Figure 6.18: Typical rise in serum enzyme activities following a myocardial infarction

 Pepsin, trypsin peptidase, lipase, amylase elastase and cellulase are used in gastrointestinal tract (GIT) disorders and chronic pancreatitis.

## Analytical Use of Enzymes

- Enzymes can be used as reagents and labels.
- In addition to measurement of serum enzyme activity for the diagnosis and management of disease, enzymes are widely used in the clinical laboratory as reagents for the estimation of serum constituents. Some examples are given in Table 6.11.
- Many enzymes have been used as the label in various immunoassays (enzyme linked immunosorption assay, ELISA) for determining the serum concentration of drugs, hormones or other compounds of interest. Commonly used label enzymes are:
  - Glucose-6-phosphate dehydrogenase
  - Alkaline phosphatase
  - β-galactosidase
  - Peroxidase.

Table 6.10: Enzymes in diagnosis of myocardial infarction and time course of plasma enzyme activity changes after myocardial infarction				
Enzyme	Abnormal activity detectable (hours)	Time for maximum rise (hours)	Time for return to normal (Days)	
CK <sub>2</sub> (MB-isoenzyme)	3-10	12-24	2-3	
AST LDH (heart specific)	6-12 8-16	24-48 48-72	4-6 7-12	